

SIXTH EDITION

ESSENTIAL ^{Best Available Copy} IMMUNOLOGY

Ivan M. Roitt

MA, DSc(Oxon), FRCPath, Hon MRCP (Lond), FRS
Professor and Head of
Departments of Immunology and Rheumatology Research
University College and Middlesex School of Medicine
University College
London W1P 9PG

BLACKWELL SCIENTIFIC PUBLICATIONS

OXFORD LONDON EDINBURGH

BOSTON PALO ALTO MELBOURNE

Exhibit 4

Best Available Copy

© 1971, 1974, 1977, 1980, 1984, 1988 by

Blackwell Scientific Publications

Editorial offices:

Osney Mead, Oxford OX2 0EL

(Orders: Tel. 0865 240201)

8 John Street, London WC1N 2ES

23 Ainslie Place, Edinburgh EH3 6AJ

3 Cambridge Center, Suite 208, Cambridge

Massachusetts 02142, USA

667 Lytton Avenue, Palo Alto, California 94301, USA

107 Barry Street, Carlton, Victoria 3053, Australia

All rights reserved. No part of this publication may be reproduced, stored in a retrieval system, or transmitted, in any form or by any means, electronic, mechanical, photocopying, recording or otherwise without the prior permission of the copyright owner

First published 1971

Reprinted 1972 (twice), 1973 (twice)

Second edition 1974, Reprinted 1975

Third edition 1977, Reprinted 1978, 1979

Fourth edition 1980, Reprinted, 1982, 1983

Fifth edition 1984

Sixth edition 1988

Reprinted 1988

Spanish editions 1972, 1975, 1978, 1982

Italian editions 1973, 1975, 1979

Portuguese editions 1973, 1976

French editions 1975, 1979

Dutch editions 1975, 1978, 1982

Japanese editions 1976, 1978, 1982, 1986

German editions 1977, 1984

Polish edition 1977

Greek edition 1978

Slovak edition 1981

Indonesian edition 1985

ELAS editions 1978, 1982, 1988

DISTRIBUTORS

USA

Year Book Medical Publishers

35 East Wacker Drive

Chicago, Illinois 60601

(Orders: Tel. 312 726-9733)

Canada

The C.V. Mosby Company

5240 Finch Avenue East,

Scarborough, Ontario

(Orders: Tel. 416-298-1588)

Australia

Blackwell Scientific Publications

(Australia) Pty Ltd

107 Barry Street

Carlton, Victoria 3053

(Orders: Tel. (03) 347 0300)

British Library

Cataloguing in Publication Data

Roitt, Ivan M.

Essential immunology. — 6th ed.

1. Immunology

I. Title

616.07'9 . QR181

ISBN 0-632-01994-8

Set by Setrite Ltd, Hong Kong

Printed and bound by

Dah Hua Printing Press Co Ltd, Hong Kong

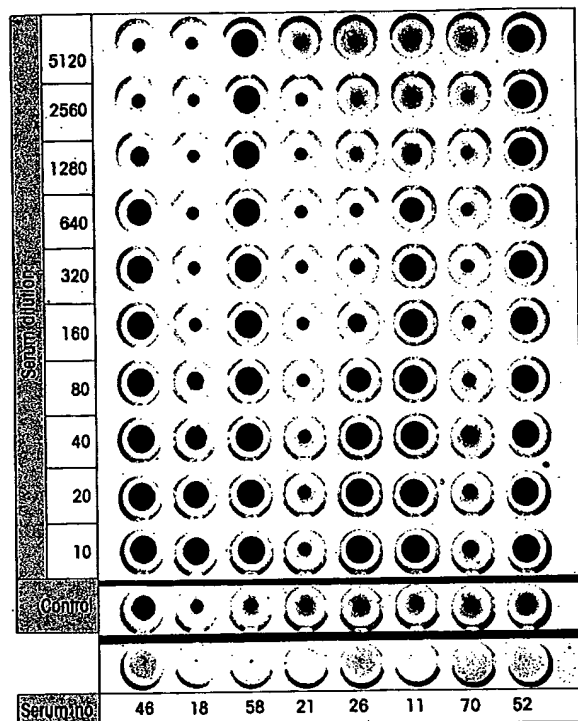


Figure 5.15. Tanned red cell haemagglutination test for thyroglobulin autoantibodies. Thyroglobulin-coated cells were added to dilutions of patients' sera. Uncoated cells were added to a 1:10 dilution of serum as a control. In a positive reaction, the cells settle as a carpet over the bottom of the cup. Because of the 'V'-shaped cross-section of these cups, in negative reactions the cells fall into the base of the 'V' forming a small easily recognizable button. The reciprocal of the highest serum dilution giving an unequivocally positive reaction is termed the titre. The titres reading from left to right are: 640, 20, >5120, neg, 40, 320, neg, >5120. The control for serum No. 46 was slightly positive and this serum should be tested again after absorption with uncoated cells.

Purification of antigens and antibodies by affinity chromatography

The principle is simple and very widely applied. Antigen or antibody is bound through its free amino groups to cyanogen-bromide-activated Sepharose particles. Insolubilized antibody, for example, can be used to pull the corresponding antigen out of solution in which it is present as one component of a complex mixture, by absorption to its surface. The uninteresting garbage is washed away and the required ligand released from the affinity absorbent by disruption of the antigen-antibody bonds by changing the pH or adding chaotropic ions such as thiocyanate (figure 5.17). Likewise, an antigen

immunosorbent can be used to absorb out an antibody from a mixture whence it can be purified by elution.

Immunoassay of antigen and antibody with labelled reagents

Antigen and antibody can be used for the detection of each other and an ingenious plethora of immunoassay techniques have been developed in which the final read-out of the reaction involves a reagent conjugated with an appropriate label.

A wide variety of labels is available

Radiolabelling with ^{131}I , or now more usually ^{125}I , is a tried and trusted technique with a very long history. Because of health hazards and the deterioration of reagents through radiation damage, other types of label have been sought. Enzymes such as peroxidase and phosphatase which give a coloured reaction product have been successfully employed particularly in the ELISA (enzyme-linked immunosorbent assay), an immunoradiometric assay for antibody and sometimes for antigen. One clever ploy for amplifying the phosphatase reaction is to use NADP as a substrate to generate NAD which now acts as a co-enzyme for a second enzyme system. Conjugation with the vitamin biotin is finding increasing favour since this can be readily detected by its reaction with enzyme-linked avidin to which it binds with ferocious specificity and affinity ($K = 10^{15}\text{M}^{-1}$). Other active approaches utilize chemiluminescent and new-style fluorescent tags.

Soluble phase immunoassays

For antibody

If a reasonable excess of labelled antigen is added to an antiserum, most of the antibodies of moderate affinity will be complexed and precipitation of the complexes followed by measurement of the label will give an estimate of the antigen binding capacity of the serum (figure 5.18). By using antibodies to different immunoglobulin classes and subclasses as the antiglobulin reagent, it is possible to determine the distribution of antibody activity among the classes. For example, addition of a radioactive antigen to human serum followed by a precipitating

Figure 5.16. Macroscopic agglutination of latex coated with human IgG by serum from a patient with rheumatoid arthritis. This contains rheumatoid factor, an autoantibody directed against determinants on IgG. (a) Normal serum, (b) patient's serum.

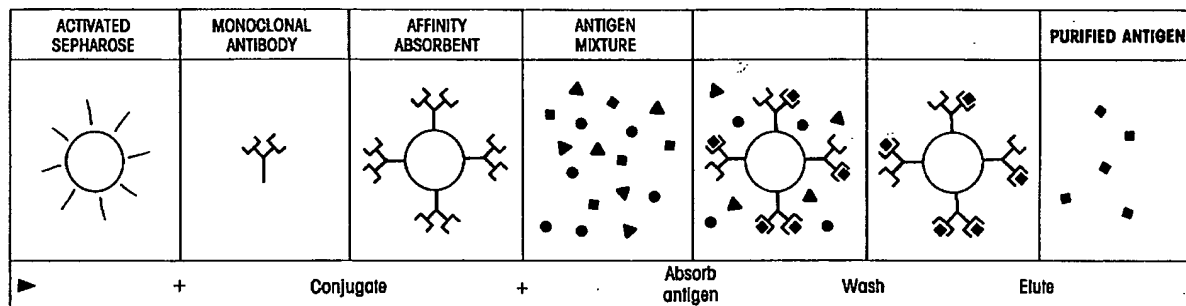
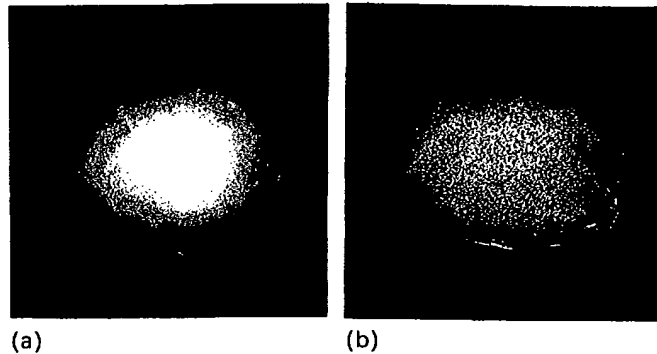
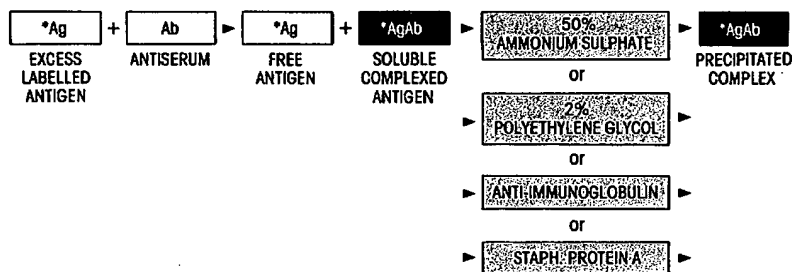


Figure 5.17. Affinity chromatography. A column is filled with Sepharose-linked antibody. The antigen mixture is poured down the column. Only the antigen binds and is released by

change in pH for example. An antigen-linked affinity column will purify antibody obviously.

Figure 5.18. Determination of antigen-combining capacity of an antiserum using antigen coupled to ^{125}I or some other label (cf. figure 5.12). The radioactivity of the precipitate provides a measure of the antigen-combining capacity.



rabbit anti-human IgA would indicate how much antigen had been bound to the serum IgA.

Classical radioimmunoassay (RIA) for antigen

The binding of radioactively labelled antigen to a limited fixed amount of antibody can be partially inhibited by addition of unlabelled antigen and the extent of this inhibition can be used as a measure of the unlabelled material added. The principle of this form of saturation analysis is explained in figure 5.19. Methods vary in the means used to separate free antigen from that bound to antibody and we have discussed the main ones already.

With the development of methods for labelling antigens to a high specific activity, very low concentrations down to the 10^{-12} g/ml level can be detected and most of the protein hormones can now be assayed with this technique. One disadvantage is that these methods cannot distinguish active protein molecules from biologically inactive fragments which still retain antigenic determinants. Other applications include the assay of carcinoembryonic antigen, hepatitis B (Australia) antigen and smaller molecules such as steroids, prostaglandins and morphine-related drugs (appropriate antibodies are raised by coupling to an immunogenic carrier).